

Synthesis and *In Vivo* Antitumor Efficacy of PEGylated Poly(L-lysine) Dendrimer–Camptothecin Conjugates

Megan E. Fox,[†] Steve Guillaudeau,[†] Jean M. J. Fréchet,^{*,†} Katherine Jerger,[‡]
Nichole Macaraeg,[‡] and Francis C. Szoka^{*,‡}

College of Chemistry, University of California, Berkeley, California 94720-1460, and
Department of Biopharmaceutical Sciences and Pharmaceutical Chemistry, University of
California, San Francisco, California 94143-0446

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Abstract: Polymer conjugates of camptothecin (CPT) have been pursued as a solution to the difficulties present in treating cancers with CPT and its derivatives. Covalent attachment of CPT to a polymer can improve solubility, increase blood circulation time, enhance tumor uptake, and significantly improve efficacy of the drug. In this report, we describe a novel polymer conjugate of CPT using a core-functionalized, symmetrically PEGylated poly(L-lysine) (PLL) dendrimer. The PEGylated dendrimer consisted of a lysine dendrimer functionalized with aspartic acid, which was used as an attachment site for poly(ethylene glycol) (PEG) and CPT. The final conjugate had a molecular weight of 40 kDa and was loaded with 4–6 wt % CPT. Polymer-bound CPT was shown to have a long blood circulation half-life of 30.9 ± 8.8 h and a tumor uptake of $4.2 \pm 2.3\%$ of the injected dose/g of tissue, compared to free CPT in which less than 1% was retained in the blood after 30 min and had a tumor accumulation of $0.29 \pm 0.04\%$ of the injected dose/g of tissue. The PEGylated PLL–CPT showed superior efficacy in murine (C26) and human colon carcinoma (HT-29) tumor models when compared with no treatment or treatment with irinotecan. In the C26 tumor model, treatment resulted in significantly prolonged survival ($P < 0.05$) for all mice treated with a single injection of PEGylated PLL–CPT. In the HT-29 tumor model, all mice treated with multiple injections of a low dose survived to the end of the study, with three mice of eight surviving tumor-free.

Keywords: Drug delivery; camptothecin; colon carcinoma; dendrimer; PEG; poly(L-lysine)

Introduction

20(S)-Camptothecin (CPT), a highly toxic alkaloid isolated from the plant *Camptotheca acuminata*, has been found to have broad antitumor activity in a variety of cancers.¹ CPT acts by inhibiting topoisomerase-I, an enzyme that stabilizes and rejoins DNA breaks during replication. However, camptothecin has never successfully been transferred to clinical

use due to poor solubility, high systemic toxicity, and decreased activity *in vivo*. In particular, at physiological pH, the lactone ring of CPT hydrolyzes to form a carboxylate that readily binds with serum albumin, which results in the removal of active CPT from the bloodstream.² Several small-molecule derivatives of CPT have been designed to improve the solubility and bioavailability of the drug, of which topotecan and irinotecan (CPT-11) are used clinically to treat certain human cancers.³ Nevertheless, topotecan and irino-

* Corresponding authors. J.M.J.F.: University of California, Department of Chemistry, 718 Latimer Hall, MC 1460, Berkeley, CA 94720-1460; e-mail, frechet@berkeley.edu; tel, (510)-643-3077; fax, (510)-643-3079. F.C.S.: Department of Biopharmaceutical Sciences and Pharmaceutical Chemistry, University of California, San Francisco, CA 94143-0446; e-mail, Szoka@cgl.ucsf.edu; tel, (415)-476-3895; fax, 415-476-2744.

[†] College of Chemistry.

[‡] Department of Biopharmaceutical Sciences and Pharmaceutical Chemistry.

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tecans still have suboptimal toxicity profiles and pharmacokinetics, and alternative methods of treatment are needed.

A method of addressing the problems associated with CPT is attachment of CPT to a polymer carrier. Polymer drug carriers have been studied extensively as a solution to many of the problems of small-molecule chemotherapeutic agents. Covalent attachment of a drug to a polymer or physical entrapment of a drug within a polymer system has been shown to increase drug solubility, improve pharmacokinetics, and decrease systemic toxicity.^{4,5} Additionally, Maeda and co-workers have shown that macromolecules passively target solid tumors through the enhanced permeation and retention (EPR) effect, resulting in higher drug concentrations in the tumor.^{6,7} Furthermore, researchers have shown that covalent modification of CPT by esterification of the alcohol at the 20-position carbon stabilized the lactone ring at physiological pH.⁸ Therefore, the esterification of CPT with a polymer at the 20-position alcohol could also prevent loss of activity associated with lactone ring-opening.

Many researchers have begun work toward a clinically viable polymer–CPT conjugate, primarily focusing on linear polymer carriers. Several conjugates have entered or progressed to phase I/II trials, including the cyclodextrin-PEG copolymer CPT conjugate IT-101 (Calando Pharmaceuticals, Pasadena, CA),^{9–12} MAG-CPT, a HPMA–CPT conjugate

(Pharmacia and Upjohn, Milan, Italy),^{13,14} CT-2106, a poly(L-glutamic acid)–CPT conjugate (Cell Therapeutics, Seattle, WA),^{15–17} and Prothecan, a PEG–CPT conjugate (Enzon, Piscataway, NY).^{18–21} IT-101 has just entered phase I clinical trials. Phase I studies of MAG-CPT showed prolonged release of CPT over 46 days,²² however, subsequent phase I/II trials resulted in excessive systemic toxicity to patients bearing tumors²³ and revealed that MAG-CPT

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showed no preferential accumulation of polymer-bound CPT within the tumors.²⁴ CT-2106 showed an improved toxicity profile compared to unconjugated CPT in phase I trials, and is progressing through phase II.²⁵ Prothecan showed promise in phase I trials with noted antitumor activity.²⁶ Despite the number of polymer–CPT conjugates in clinical trials, none have been approved for treatment yet.

PEGylated dendrimers are being pursued for drug delivery applications due to their improved *in vivo* pharmacokinetic properties, decreased toxicity, and enhanced solubility compared to non-PEGylated dendrimers and are particularly promising for delivery of chemotherapeutic agents.^{27–30} In contrast to conventional polymers, dendrimers are synthesized in a controlled, stepwise manner, resulting in a well-defined, branched structure with low polydispersity.³¹ Furthermore, the architecture, type and number of functional handles can be easily controlled. Researchers have PEGylated poly(amidoamine) (PAMAM),^{32,33} poly(L-lysine) (PLL),^{34–37}

and melamine³⁸ dendrimers for different biological applications. Our previous work using PEGylated dendrimers explored asymmetrically and symmetrically PEGylated polyester (PE) dendrimers based on bis(2-hydroxymethyl) propionic acid.^{39–42} A conjugate of doxorubicin (DOX) and an eight-arm, 40 kDa asymmetrically PEGylated PE dendrimer was synthesized, and studied in tumor-bearing mice. The study revealed the PEGylated dendrimer–DOX conjugate was remarkably effective at delivering DOX to the tumor, with a single, nontoxic dose resulting in complete remission of the tumor in all mice tested.⁴¹ The efficacy of this carrier was thought to be due to the core–shell architecture that results from PEGylating the periphery of a globular dendrimer. Drug was covalently attached to the low molecular weight dendrimer core and surrounded by a “shell” of long, solubilizing PEG chains which radiate from the core. Guillaudeu and researchers reported a simple and rapid method for synthesizing symmetrically PEGylated PE dendrimers that retains the core–shell architecture while incorporating multiple functional handles at the dendrimer core.⁴²

As prior work has shown the potential of PEGylated dendrimer drug carriers to significantly improve the efficacy of a chemotherapeutic drug, we have synthesized and evaluated a novel PEGylated PLL–CPT conjugate for cancer treatment. Two generation two lysine dendrons were constructed around a diaminopropane core. The dendrimer periphery was modified with aspartic acid to incorporate two

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functional handles: a carboxylic acid and an amine. The amine of the aspartic acid was PEGylated with 5 kDa PEG chains, and the carboxylate esterified with glycine-20(*O*)-CPT prodrug. The resulting PEGylated PLL–CPT conjugate had eight arms and a molecular weight of 40 kDa, and it contained 6 wt % CPT. Herein is presented the synthesis, *in vitro* and *in vivo* analysis, and chemotherapeutic effectiveness of this new polymer–CPT conjugate.

Experimental Section

Reagents. Chemicals were purchased from Aldrich and were used without further purification. ONp-activated, Boc-protected lysine was purchased from Bachem (Bubendorf, Switzerland). Boc-protected, Bn-ester-protected aspartic acid was purchased from Novabiochem (Darmstadt, Germany). ONp-activated PEG5000 (PEG5k) was purchased from Laysan Bio, Inc. (Arab, AL).

For intravenous injection, polymer–CPT conjugates were dissolved in $\text{Mg}^{2+}/\text{Ca}^{2+}$ -free PBS, and reported doses were given as drug equivalents of polymer. CPT was suspended in a solution of 10% Tween 80/0.9% NaCl (9:1) and sonicated for 20 min to break up aggregates. CPT-11 was dissolved in a solution of 45 mg/mL D-sorbitol and 0.9 mg/mL lactic acid and heated at 50 °C for 20 min. All solutions were made just prior to injection.

Synthesis of Camptothecin-Loaded PEGylated Poly(L-lysine) Dendrimers. The scheme for the synthesis of both glycine-linked PEGylated PLL–CPT (**11a**) and β -alanine-linked PEGylated PLL–CPT (**11b**) is shown in Scheme 1. Briefly, Boc-protected L-lysine, activated with *p*-nitrophenol, was reacted with diaminopropane in DMF. The first generation dendrimer (**3**) was purified by precipitation into a 0.2 M NaOH solution and collection of the solids via filtration through filter paper. After removal of the Boc-protecting groups in a mixture of TFA and DCM, **4** was made in a similar manner. The aspartic acid periphery was added by reacting the deprotected amines of **4** with Boc-Asp(Bn)-OH (**5**) in DMF with triethylamine. To PEGylate the Boc-deprotected amines of dendrimer **6**, the TFA-dendrimer salt was combined with triethylamine and ONp-activated PEG5k (**7**) in DMF at 30 °C. Unreacted PEG5k was capped with piperidine. To remove the benzyl ester protecting groups, **8** was stirred with 4% acetic acid in methanol and ~10% by weight palladium-on-carbon catalyst.

Boc-Gly-CPT and Boc- β -Ala-CPT were synthesized following a previously described procedure.²⁰ After removal of the Boc-protecting groups with TFA, the CPT prodrugs were conjugated to the free carboxylates of **9** using HBTU in the presence of DIPEA and PCP in DMF. CPT content was quantified by UV absorbance at 365 nm in chloroform ($\epsilon = 21,856 \text{ M}^{-1} \text{ cm}^{-1}$). Additional details of polymer and small molecule characterization can be found in the Supporting Information.

Camptothecin Hydrolysis Studies. To quantify the hydrolysis of CPT from the dendritic polymer carrier in various buffers, **11a** or **11b** was dissolved in either PBS without

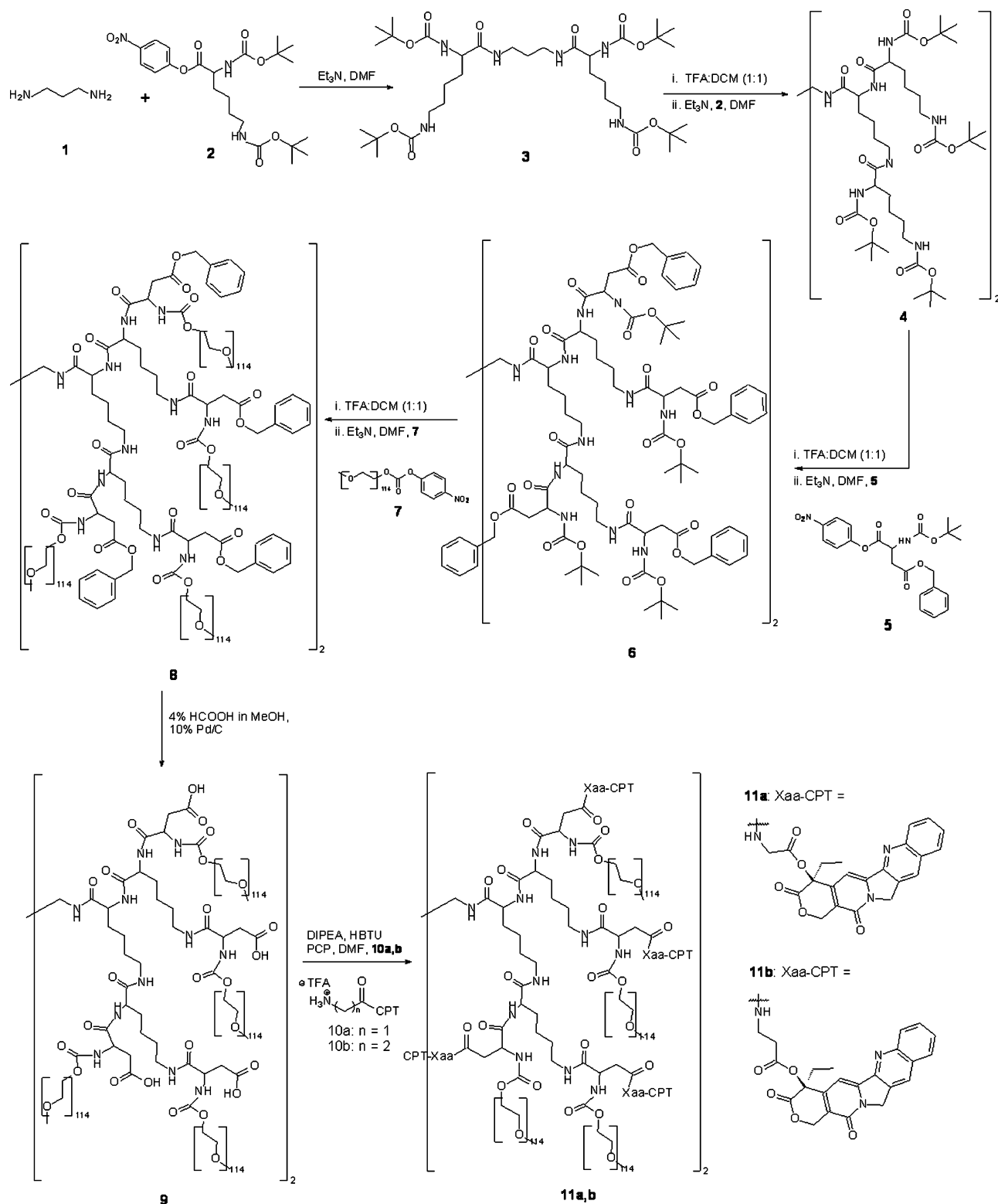
$\text{Mg}^{2+}/\text{Ca}^{2+}$ (pH 7.4), acetate buffer (30 mM with 70 mM NaNO_3 , pH 5.0), or reconstituted human plasma (adjusted to pH 7.4 with HEPES buffer (1 M HEPES, 13.5 g/L NaOH, 9 g/L NaCl)) at a concentration of 10 mg polymer/mL and incubated in the dark at 37 °C. At each time point, a 50 μL aliquot was combined with 250 μL of 90% MeOH/10% AcOH and 200 μL of chloroform and briefly vortexed. The sample was centrifuged at 12,000 rpm for 15 min, and the supernatant injected onto a RP-HPLC for analysis. The RP-HPLC system consisted of a C-8 Zorbax Exlipse XDB-C8 $4.6 \times 150 \text{ mm}$, 5 μm column (Agilent, LOCATION) in line with a Waters 996 photodiode array detector and a Waters 2414 refractive index detector, using gradient elution with distilled water (with 0.05% TFA) and acetonitrile (with 0.05% TFA).

Tumor and Animal Models. C26 murine colon carcinoma cells obtained from the University of California, San Francisco, cell culture facility were cultured in RPMI medium 1640 containing 10% FBS. HT-29 human colon carcinoma cells were acquired from Dr. B. Mark Evers at the University of Texas and were cultured in DMEM-1 with Glutamax containing 10% FBS. Female Balb/C mice (16–20 g) and female athymic *nu/nu* mice (16–20 g) were obtained from Simonsen Laboratories, Inc. (Gilroy, CA). All animal maintenance and experiments performed were in compliance with National Institutes of Health guidelines for animal research under a protocol approved by the Committee on Animal Research at the University of California, San Francisco.

Cytotoxicity Studies in Cells. Cytotoxicity studies were completed as previously described.⁴¹ Briefly, cells (C26 or HT-29) were incubated in medium (C26: RPMI 1650 with 10% FBS, 1% penicillin/streptomycin, 1% Glutamax; HT-29: DMEM-1 with Glutamax and 10% FBS) containing varying concentrations of CPT, **11a**, or **11b**. After a 72 h incubation, the medium was replaced with medium containing thiazolyl blue tetrazolium bromide (0.83 mg/mL) and incubated an additional 3 h. The medium was replaced with dimethyl sulfoxide (200 μL) and 0.1 M glycine buffer (25 μL). Optical densities were measured at 570 nm by a SpectraMAX 190 microplate reader with Softmax Pro software (Molecular Devices, Sunnyvale, CA). IC_{50} values were obtained from sigmoidal fits of the data using Origin 7 SR4 8.0552 software (OriginLab, Northampton, MA).

Acute Toxicity Studies in Healthy Mice. To determine the *in vivo* maximum tolerated dose for PEGylated PLL–CPT, the conjugate was administered intravenously (iv) to Balb/C mice in doses of 0 (PBS), 8, 16, 32, 48, and 64 mg CPT equivalents/kg mouse (3 mice per treatment group). The weights and general health of each mouse were monitored. When a mouse showed severe signs of toxicity, which included weight loss greater than 25% of the initial weight, lethargy, a hunched back, and ruffled fur, the mouse was sacrificed. The mice were monitored for 30 days.

Pharmacokinetic and Biodistribution Studies in C26-Tumored Balb/C Mice. Sixteen to twenty-five gram female Balb/C mice were injected in the right hind flank with 3 \times

Scheme 1. Synthesis of Glycine-Linked and β -Alanine-Linked PEGylated PLL–CPT Conjugates

10^5 C26 cells. Nineteen days after tumor inoculation, mice were distributed randomly among four groups. Mice were injected iv (three mice per group) either with **11a** (10 mg CPT equiv/kg) or with CPT (10 mg/kg) in $\sim 200 \mu\text{L}$ solution. The mice injected with polymer were sacrificed at 90 min, 360 min, or 2880 min postinjection. Additional time points for the blood concentration were collected by submandibular

bleeds. The mice injected with CPT were sacrificed at 1440 min. At the time of sacrifice, the blood (collected by heart puncture), lungs, liver, spleen, intestines, stomach, kidney, muscle, and tumor were collected for analyses. Organs were weighed, and 200–300 mg of the collected organs were homogenized with zirconium beads and 1 mL of acidified isopropyl alcohol (0.075 M HCl, 90% IPA). The samples

then incubated at 4 °C for 24 h. Serum was collected using Microtainer serum separator vials and processed in the same manner as the organs. All samples were then placed in a –80 °C freezer until measurements could be made. At measurement time, samples were thawed and briefly vortexed. One hundred twenty-five microliters of homogenate was combined with 875 μ L of acidified IPA, homogenized a second time, and centrifuged for 8 min at 14,000 rpm. Two hundred fifty microliters of supernatant was combined with 750 μ L of acidified IPA for fluorescence measurements. Sample fluorescence was measured on a Fluorolog fluorimeter (HORIBA Jobin Yvon, Edison, NY), with an excitation of CPT at 372 nm and collection of fluorescence at 424 nm. Calibration curves were made from organ samples collected from an untreated mouse. The pharmacokinetic parameters of **11a** were calculated assuming the two-compartment model.⁴³

Antitumor Efficacy Studies. *C26 Murine Colon Carcinoma Model.* While under anesthesia, female Balb/C mice were shaved, and C26 cells (3×10^5 cells in 50 μ L) were injected subcutaneously on the right hind flank. At eight days post-tumor implantation, mice were randomly distributed into treatment groups of 5–10 mice and treated intravenously (iv) (tail vein) as described in the text. All doses were administered in approximately 200 μ L of solution. Animals were weighed and tumors measured every other day. The tumor volume was determined by measuring the tumor in three dimensions with calipers and calculated using the formula tumor volume = length \times width \times height. Mice were removed from the study when (1) a mouse lost 15% of its body weight from the first day of treatment, (2) any tumor dimension was >20 mm, or (3) the mouse was found dead. The study was continued until day 60 post-tumor inoculation.

HT-29 Tumor Model. Female athymic *nu/nu* mice were injected subcutaneously on the right hind flank with HT-29 cells (1×10^6 cells/mL in 50 μ L). Twelve days later, mice were randomized into groups of eight animals per group and treated iv (tail vein) as described in the text. The tumor volume was calculated using the following formula tumor volume = (length \times width²)/2. The mice were followed until day 70 post-tumor inoculation.

Statistical Analyses. The tumor growth delay (TGD) was calculated from the equation $TGD = (T - C)/C \times 100$, where *T* is the mean time for the tumor volume of a treatment group to reach a designated volume and *C* is the mean time of the control group to reach the designated volume (500 mm³ for the C26 tumor model, 400 mm³ for the HT-29 tumor model). Mice that were removed from the study from treatment-related weight loss before the end of the study were not included in the statistical analysis. Mice that did not reach the designated tumor volume but were alive at the end of the study were assigned the last day of the study as the time

to tumor volume or day of survival. An independent Student's *t* test was used to analyze the TGD and survival data for all animals. All statistical analyses were performed using MedCalc 8.2.1.0 for Windows (MedCalc Software, Mariakerke, Belgium).

Results

Preparation of PEGylated PLL–CPT Conjugates. The synthesis of the PEGylated PLL dendrimer is shown in Scheme 1. Synthesis of the unPEGylated, aspartic acid-terminated lysine dendrimer is stepwise and results in a complete, monodisperse molecule. The final molecular weight by SEC was measured as ~35 kDa, although the theoretical molecular weight was 40 kDa. As previously reported, SEC underestimates the hydrodynamic volume, and thus molecular weight, of PEGylated dendrimers due to the branched nature of the polymer.³⁹ The final polydispersity of the PEGylated PLL dendrimers was ~1.1, which was essentially the polydispersity of the 5000 Da PEG chains.

Based on reports in the literature, glycine was chosen as a linker between the PEGylated PLL dendrimer and CPT because of its favorable release profile and improved efficacy in polymer–CPT systems compared to other CPT linkers.^{17,20} The second linker, β -alanine, was chosen for use as a nonreleasable polymer–CPT control, as β -alanine-linked CPT was shown to have a much slower release from polymer compared to glycine-linked CPT.¹⁷ The CPT conjugate **11b** was made to test the requirement of drug release on polymer–CPT efficacy. After synthesis of the linker–CPT prodrugs, conjugation of the prodrug was achieved through standard protein coupling chemistry using HBTU. The theoretical maximum weight loading of CPT on the PEGylated PLL dendrimer is 6.5 wt %. Weight-loadings of 6% and 4.5% were achieved for **11a** and **11b**, respectively.

In Vitro Hydrolysis of CPT from the PEGylated PLL Dendrimer. To mimic the potential *in vivo* release of CPT, hydrolysis of CPT from the PEGylated PLL dendrimer was monitored in three physiologically relevant solutions: PBS (pH 7.4), acetate buffer (pH 5.0), and human plasma (buffered to pH 7.4 with HEPES buffer) (Figure 1). We expected that free CPT would be released via hydrolysis of the ester bond linking the drug to the amino acid linker. This was verified by analyzing a sample of free CPT, which corresponded to the small molecule released from each polymer (data not shown).

Surprisingly, we discovered the hydrolysis of CPT from **11a** and **11b** was the same in both PBS buffer and human plasma (Figure 1). Many researchers have chosen amino acid linkers for CPT because it is thought that CPT release could be achieved by enzymatic cleavage, and in fact, many observe faster release rates in the presence of cathepsin B or human plasma.^{13,18,20,44} However, we observed similar

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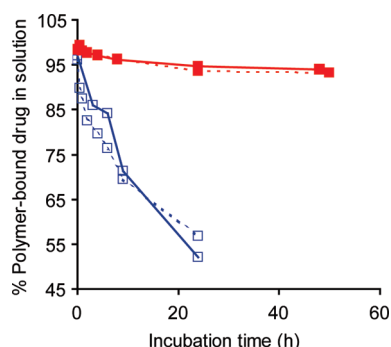


Figure 1. Hydrolysis of **11a** and **11b** in physiological solutions, given as the % of the polymer-bound CPT in solution as a function over time. **11a** (blue open squares) and **11b** (red filled squares) were incubated in either PBS without Mg^{2+}/Ca^{2+} salts, pH 7.4 (—) or reconstituted human plasma, buffered to pH 7.4 with HEPES buffer (---).

Table 1. IC_{50} Values of Polymer–CPT Conjugates and Free CPT in Cells^a

polymer/drug	C26 (nM)	HT-29 (nM)
11a	67.6 ± 6.5	522 ± 95
11b	1030 ± 80	NA
CPT	34.4 ± 7.0	38.3 ± 5.0

^a Values determined by the MTT assay.

release profiles of CPT in the presence and absence of enzymes. In both buffers, the hydrolysis of CPT from **11a** had a half-life of 20 h, and hydrolysis of CPT from **11b** had half-life of ~200 h. The release of CPT in acetate buffer was less than 2% after 24 h for both polymers (data not shown). The faster hydrolysis of CPT at pH 7.4 compared to hydrolysis at pH 5.0 was expected, as ester bonds are less stable at higher pH.

In Vitro Toxicity of PEGylated PLL–CPT Conjugates.

The cytotoxicity of CPT and each polymer–CPT conjugate in C26 cells and HT-29 cells is given in Table 1. In the C26 line, conjugation of CPT via the glycine linker (**11a**) reduced the toxicity of CPT to the cells by approximately 2-fold. Conjugation of CPT via the β -alanine linker (**11b**) reduced the toxicity by nearly two orders of magnitude. This decrease in toxicity of **11b** reflected the slower hydrolysis of CPT from the polymer. In the HT-29 cell line, the toxicity of **11a** was reduced by more than one order of magnitude, compared to free CPT.

Pharmacokinetics and Biodistribution in Tumor Mice. The pharmacokinetics and body distribution of **11a** and CPT were determined in C26-tumor-bearing female Balb/C mice (Figure 2). Less than 1% of the injected dose of free CPT was retained in the blood after 30 min. In contrast, conjugation of CPT to the PEGylated PLL dendrimer significantly prolonged blood circulation, resulting in a blood circulation half-life of 30.9 ± 8.8 h. Furthermore, area-under-the-curve of **11a** was calculated to be $460 \pm 140\%$ of the injected dose-h/gram of tissue (% ID-h/g of tissue). For polymer–drug conjugates, a large area-under-the-curve is desirable because increased systemic exposure results in

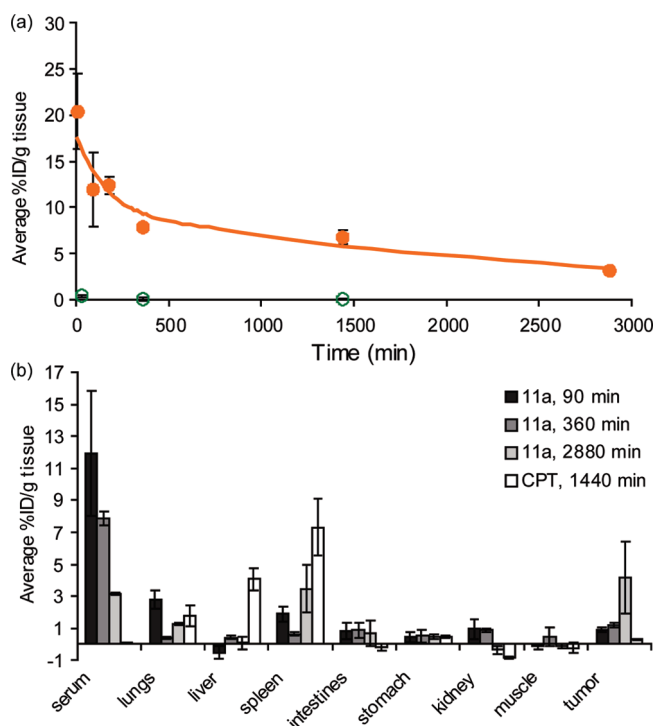


Figure 2. (a) Blood circulation profile of **11a** (●) and free CPT (○) given as the average % ID/g of blood over time. The solid line represents the calculated blood curve of **11a**, using the two-compartment model. (b) Biodistribution of **11a** and free CPT given as the average % ID/g of tissue collected at 90 min, 6 h, and 48 h for **11a** and collected at 24 h for free CPT. Negative values for the average % ID/g of tissue are a result of measurements at or near the background fluorescence of CPT.

increased tumor exposure to the drug, and can result in high drug accumulation in the tumor.

Given the long blood circulation half-life, the biodistribution of **11a** in tumor-bearing mice was collected at three time points postinjection, with the last time point at 48 h postinjection. However, only a 24 h postinjection time point was collected for free CPT (Figure 2b). As can be seen, free CPT accumulated in the lungs, liver, and spleen, with accumulations of $1.75 \pm 0.65\%$ ID/g of tissue, $4.07 \pm 0.72\%$ ID/g of tissue, and $7.33 \pm 1.77\%$ ID/g of tissue, respectively. Accumulation of free CPT in the tumor tissue was much less, with only an accumulation of $0.29 \pm 0.04\%$ ID/g of tissue. Conjugation of CPT to the PEGylated PLL dendrimer decreased accumulation of CPT in the lungs, liver, and spleen, and significantly improved accumulation of CPT in the tumor. After 48 h, the accumulation of CPT in the tumor was $4.2 \pm 2.3\%$ ID/g of tissue. In fact, the accumulation of CPT in the tumor from polymer-bound CPT was significantly improved after only 6 h, when compared with the accumulation of free CPT after 24 h.

The biodistribution of both free CPT and the polymer–CPT conjugate was determined using the fluorescence of CPT at 424 nm. While the pharmacokinetics and biodistribution of polymer-bound CPT could be that of free CPT and **11a**, we

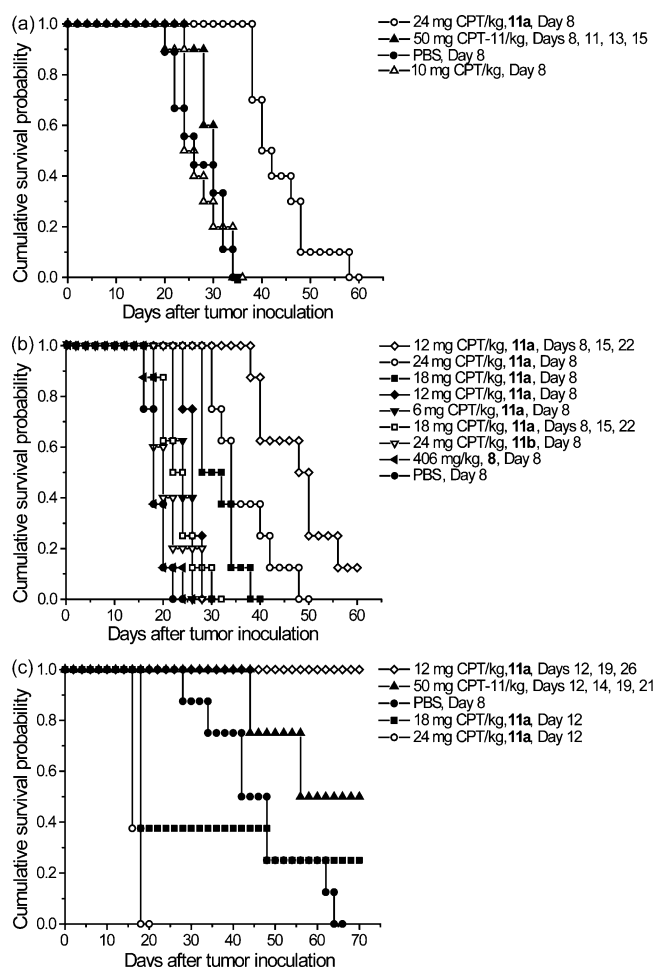


Figure 3. (a) Antitumor efficacy in the C26 tumor model, free drug control experiment. (b) Antitumor efficacy in the C26 tumor model, dose escalation and multiple doses schedule. (c) Antitumor activity in the HT-29 tumor model.

feel confident that we are observing primarily polymer-bound CPT given the rapid removal of free CPT from the body. Furthermore, we note that negative values for tissue accumulation were a result of measurements that were at or below the background level of fluorescence signal at 424 nm. Thus, we concluded very little CPT was present in these measurements.

Efficacy of PEGylated PLL–CPT in C26-Tumored Mice. CPT derivatives are used clinically to treat colorectal cancers. We chose to test our PEGylated PLL–CPT conjugates in the murine colon carcinoma model C26. After determining the maximum tolerated dose in healthy Balb/C mice to be between 32 mg CPT/kg and 48 mg CPT/kg (data not shown), two chemotherapy experiments were performed in C26-tumored Balb/C mice, and are detailed in Figure 3 and Table 2. As CPT is not typically tested in the C26 tumor model, an initial study was completed to compare **11a** against free drug controls. In the drug controls study, a single, high dose of **11a** was tested against free CPT and irinotecan at either a single or multiple injections (Table 2a, Figure 3a). The dose of 24 mg CPT/kg was tested, as it was 75% of the

highest tolerated dose in healthy mice. A single dose of 10 mg CPT/kg was chosen based on the toxicity of CPT in the Balb/C mouse strain. Irinotecan was given at a dose of 50 mg CPT-11/kg for a total of four doses, and was included as a second free drug control. While it was found that neither CPT nor irinotecan had any significant effect on the survival of the C26-tumored mice, irinotecan, given twice weekly for four total doses, significantly delayed tumor growth (18% TGD) compared to no treatment ($P \leq 0.01$). Despite no significant survival response from either free drug, mice treated with **11a** showed significantly increased survival ($P \leq 0.0001$) and improved TGD (72%) compared to both PBS and irinotecan.

Given the promising results of the first C26 chemotherapy study, a second chemotherapy study in C26-tumored mice was completed, looking at single-injection dose escalation (6, 12, 18, and 24 mg CPT/kg) and multiple-injection schedules (12 mg CPT/kg and 18 mg CPT/kg, given once weekly for a total of 3 injections) of **11a**. A treatment of 24 mg of CPT/kg of **11b** was given as a control in which CPT was “nonreleasable”. PEGylated dendrimer **8** was also treated at a dose of 406 mg of polymer/kg, equivalent polymer to a dose of 24 mg of CPT/kg, as a polymer-only control. Treatment with **11a** proved to be remarkably effective at prolonging tumor growth delay and survival of the mice. All groups treated with a single injection of **11a**, even down to a dose of 6 mg of CPT/kg, showed significant tumor growth delay and increased survival ($P \leq 0.0001$). As for treatment with multiple injections of **11a**, all mice treated with a dose of 18 mg of CPT/kg suffered severe weight loss after the second and third injections, and were removed from the study as treatment-related deaths. However, mice treated with three weekly injections of 12 mg of CPT/kg had the greatest TGD at 155% and longest median survival time. The median survival time was 49 days, with one mouse being tumor-free at day 60. Additionally, no significant weight loss was observed. As expected, treatment with either **11b** or **8** resulted in TGD and survival similar to no treatment (PBS).

Efficacy of **11a in HT-29-Tumored Mice.** Several researchers have tested the efficacy of polymer–CPT conjugates in the HT-29 human colon carcinoma model.^{12–14,19,21} Likewise, we tested the efficacy of **11a** in HT-29-tumored *nu/nu* athymic female mice (Figure 3 and Table 2). Mice were treated with either a single injection of 24 mg of CPT/kg or 18 mg of CPT/kg or three weekly injections of 12 mg of CPT/kg. Irinotecan, given at 50 mg of CPT-11/kg twice weekly for four injections, was used as a positive control.⁴⁵ The irinotecan schedule was found to significantly delay tumor growth (60% TGD) and prolong survival compared to no treatment ($P \leq 0.05$), and four mice survived to the end of the study (day 70). Treatment with **11a** with the once weekly injection of 12 mg of CPT/kg, however, had a significantly prolonged TGD of 122% compared to both PBS

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Table 2. Antitumor Activity of **11a** in the C26 and HT-29 Colon Carcinoma Models^a

experiment/ treatment	<i>n</i>	dose (mg/kg)	no. of injections	treatment days	mean TGD (%)	median survival time (days)	TRD	TFS	LTS
(a) C26: Free Drug Controls									
PBS	9		1	8		26	0	0	0
CPT	10	10	1	8	<5	25	0	0	0
CPT-11	10	50	4	8, 11, 13, 15	18 ^c	30	0	0	0
11a	10	24	1	8	72 ^{b,d}	41 ^{b,d}	0	0	0
(b) C26: Dose Escalation and Multiple Doses									
PBS	8		1	8		18	0	0	0
8	8	406	1	8	1.5	18	0	0	0
11b	5	24	1	8	10	20	0	0	0
11a	8	24	1	8	89 ^b	34 ^b	0	0	0
11a	8	18	1	8	59 ^b	30 ^b	0	0	0
11a	8	12	1	8	47 ^b	26 ^b	0	0	0
11a	8	6	1	8	39 ^b	24 ^c	0	0	0
11a	8	18	3	8, 15, 22	NA	23	8	0	0
11a	8	12	3	8, 15, 22	155 ^b	49 ^b	0	1	1
(c) HT-29: Dose Escalation, Multiple Doses, and Free Drug Controls									
PBS	8		1	8		45	0	0	0
CPT-11	8	50	4	12, 14, 19, 21	60 ^c	63 ^c	0	0	4
11a	8	24	1	12	NA	16	8	0	0
11a	8	18	1	12	97 ^c	18	5	0	2
11a	8	12	3	12, 19, 26	122 ^{b,e}	70 ^{b,e}	0	3	8

^a See *Experimental Section* for experimental details and statistical methods. Dose given in drug or drug equivalents, except for **8**, which is given as dose of polymer. TGD, tumor growth delay, calculated from time of growth to 500 mm³ (C26) or 400 mm³ (HT-29); TRD, treatment-related death; TFS, tumor-free survivors; LTS, long term survivors. ^b Compared to PBS, $P \leq 0.0001$. ^c Compared to PBS, $P \leq 0.05$. ^d Compared to CPT-11, $P \leq 0.0001$. ^e Compared to CPT-11, $P \leq 0.05$.

($P \leq 0.0001$) and irinotecan ($P \leq 0.05$), and all mice survived to the end of the study. Three of eight mice were tumor-free. Mice treated with either 24 mg of CPT/kg or 18 mg of CPT/kg of **11a** showed excessive weight loss within 10 days of treatment and were removed from the study as treatment-related deaths. Of the three mice that survived the 18 mg of CPT/kg of **11a** treatment, tumor growth was significantly delayed compared to PBS ($P \leq 0.01$).

Discussion

Polymer drug carriers are studied for their potential to reduce drug toxicity, improve bioavailability, increase drug solubility and enhance targeting.⁴ Anticancer drugs covalently attached to polymer via a releasable linkage more effectively target tumor tissue than drug alone through the EPR effect.⁷ With a careful design of the drug linkage, the drug can remain attached to the polymer while circulating in the blood compartment, and be released after the polymer drug carrier is taken up into the tumor tissue.

Dendrimers have been explored for drug delivery applications due to their well-defined size, shape, and controlled exterior functionality.^{27–29,46–49} Our group has extensively studied dendritic polymer architectures for the

delivery of anticancer drugs to tumors.^{40–42,50–53} In particular, we had remarkable success treating the C26 colon carcinoma model with a 40 kDa asymmetrically PEGylated polyester dendrimer–DOX conjugate, termed the “bowtie”.⁴¹ However, due to its asymmetric nature, the synthesis of this drug carrier was labor intensive, and

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a simpler symmetric, core-functional PEGylated dendrimer was designed in which the favorable pharmacokinetics and multifunctionality of the “bowtie”-type dendrimers were retained.⁴² In the present study, we applied the symmetric, core-functional design to PEGylated PLL dendrimers and covalently attached CPT. The PEGylated PLL dendrimer–CPT conjugates were analyzed for their *in vitro* and *in vivo* properties, and their antitumor efficacy in the C26 murine and HT-29 human colon carcinoma models studied.

The synthesis of PEGylated PLL dendrimers following the core-functional design is a simple stepwise procedure, resulting in a well-characterized drug carrier with low polydispersity. One of the advantages of this method is the ease in which multiple functionalities are incorporated into the periphery of the dendrimer using simple amino acids. Aspartic acid was chosen to add multifunctionality to the lysine dendrimer. Furthermore, the stepwise, column-free procedure for making peptide dendrimers allows for rapid scale-up of the synthesis, and large quantities of the PEGylated PLL drug carrier can be made easily.

To covalently attach camptothecin to a polymer backbone, multiple linkages based on amino acids and small peptides have been studied.^{17,18,20,44} Of those studied, polymer–camptothecin conjugates using a linker consisting of a single glycine residue are frequently found the most efficacious due to the favorable stability and release profile of glycine–CPT conjugates.^{17,20} Based on these prior results, we chose to examine the efficacy of our PEGylated PLL dendrimers in delivering CPT using a glycine linkage between CPT and the aspartic acid carboxylate. The amino acid linker β -alanine was chosen to verify the requirement of a releasable drug linkage in the efficacy of our drug carrier. A previous study indicated that CPT attached to a polymer backbone via β -alanine would have a much slower release rate and lower efficacy.¹⁷ This was verified by hydrolysis studies, which showed the release of CPT from β -alanine linkage was an order of magnitude slower than release of CPT from the glycine linkage. Surprisingly, hydrolysis studies of **11a** and **11b** in PBS and plasma revealed no difference in CPT release in the presence of plasma proteins and enzymes. This suggested to us that the release of CPT from the PEGylated PLL dendrimer core occurred primarily by hydrolysis. It is possible that the core–shell architecture of the PEGylated dendrimers prevents enzymes and plasma proteins from accessing the drug linkage near the core of the PEGylated dendrimer. Similarly, prior work from our group observed nearly identical degradation rates in plasma and PBS buffer observed for PEGylated polyester dendrimers.⁴²

To the best of our knowledge, only a few reports exist testing CPT or its derivatives in a subcutaneously implanted C26 tumor model.^{54,55} Therefore, we conducted an initial efficacy study in subcutaneously-C26-tumored Balb/C mice

using a single high dose of CPT, a single high dose of **11a**, and 4 injections of CPT-11, given every two days. Overall, we found neither CPT nor CPT-11 to be effective in prolonging the survival of mice when using these schedules. However, a single, high dose of our **11a** significantly prolonged mouse survival time and delayed tumor growth. This improved efficacy of polymer-bound CPT may be a result of not only the increased blood circulation time but increased tumor accumulation of CPT. Furthermore, a second study in C26-tumored Balb/C mice with **11a** showed mice treated with a single injection as low as 6 mg CPT/kg had significantly prolonged survival and TGD when compared to the control mice. As expected, a dose-dependent response was observed, with mice treated with higher doses having higher median survival times and higher TGD. However, we did observe that mice treated at higher doses (18 mg CPT/kg and 24 mg CPT/kg) showed some weight loss in the week following treatment. This weight loss proved to be too extensive when a multiple-injection schedule was used for the dose of 18 mg CPT/kg. All mice were removed from the study due to excessive weight loss from **11a** treatment. On the other hand, a multiple-injection schedule at a lower dose of **11a**, 12 mg CPT/kg, proved to be the most effective out of all treatment groups at prolonging survival time and delaying tumor growth without any significant weight loss from treatment.

A multiple-injection treatment schedule at a dose of 12 mg of CPT/kg of **11a** was also found to be the most effective in delaying tumor growth and prolonging survival in the HT-29 tumor model. At the end of the study, all mice had survived with an average tumor volume <300 mm³ in mice bearing tumors, and three mice were tumor-free. Additionally, we observed that a multiple-injection treatment of irinotecan had significant tumor delay and prolonged survival in the HT-29 tumor model, as predicted.⁴⁵ However, a multiple-injection treatment of **11a** at the dose of 12 mg of CPT/kg significantly improved tumor delay and survival over CPT-11. This result is comparable to results obtained by Schluep and co-workers with their cyclodextrin–CPT conjugate IT-101 using the approximately the same treatment schedule and dose.¹² However, our polymer is of a lower molecular weight than IT-101, which could result in less long-term accumulation. Unfortunately, many of the mice treated with a single high dose of either 18 or 24 mg of CPT/kg of **11a** observed excessive weight loss and were removed from the study as treatment-related deaths. The HT-29 model is a human tumor model and must be tested in athymic nude mice, thus we anticipated that the nude mice would be more sensitive to **11a** than the Balb/C mice, and we observed greater toxic effects at equivalent doses.

Taken together, these results indicate that the PEGylated PLL dendrimer carrier effectively improved the bioavail-

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ability, solubility, and efficacy of the anticancer drug CPT. Furthermore, the glycine–CPT conjugate, **11a**, was effective against the colon carcinoma models C26 and HT-29 by prolonging survival time and significantly slowing tumor growth. As seen in previous studies, the PEGylated dendrimer polymer drug carrier has proven to effectively prolong blood circulation time, increase tumor accumulation, and protect its drug cargo from enzymatic access. To the best of our knowledge, this is the first paper reporting the synthesis and *in vivo* evaluation of a PEGylated poly(L-lysine) dendrimer–drug conjugate. We are continuing to test the PEGylated PLL dendrimer–CPT in additional efficacy studies.

Abbreviations Used

Boc, *N*-*tert*-butoxycarbonyl; Bn, benzyl; CPT, camptothecin; CPT-11, irinotecan; DCM, dichloromethane; DIPEA, *N,N*-diisopropylethylamine; DMAP, 4-dimethylaminopyridine; DMF, dimethylformamide; HBTU, *O*-benzotriazole-

N,N,N',N'-tetramethyl-uronium-hexafluoro-phosphate; IPA, isopropyl alcohol; LTS, long-term survivors; ONp, *p*-nitrophenyl ester; PCP, pentachlorophenol; PEG, polyethylene glycol; PLL, poly(L-lysine); RP-HPLC, reverse-phase high performance liquid chromatography; TFA, trifluoroacetic acid; TFS, tumor-free survivors; TGD, tumor growth delay; TRD, treatment-related death.

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Supporting Information Available: Detailed synthetic methods and characterization for the PEGylated PLL–CPT conjugates and intermediates. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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